

Microarray Technique for Studying Genetic Diversity in Saudi Sickle Cell Patients

Awwad Alenezy, MD* Dalal S.AlShaya, Ph.D** Hayat Alafari, Ph.D** Amna Alotiby, Ph.D*** Talat Bukhari, Ph.D*** Abdulkarim S. BinShaya, Ph.D**** Hisham Waggiallah, Ph.D**** Wael Alturaiki, Ph.D***** Yasser Alnaam, Ph.D***** Arwa F. Alanazi, BchD***** Amani F. Alanazi, BRT***** Faris Q.B.Alenzi, Ph.D****

ABSTRACT

Introduction: Sickle cell anemia is one of the most common heritable hematologic diseases affecting humans. Approximately 3 million individuals had genetic blood diseases in Saudi Arabia, of whom 30% lived in Dammam Region. The aim of this study was to conduct complete gene survey studies using microarray technology.

Material and Methods: Blood samples from 90 unrelated sickle cell disease patients were obtained from the KCUH, Riyadh between from January 2017 and to June 2020. In this study, linkage disequilibrium has been determined between single nucleotides polymorphism loci in the same region of beta globin gene to identify which of them had a role for the unique variable appearance of the disease affect. To achieve such goal, the Haploview program was used.

Results: The obtained results revealed the region from 5246694 to 5251625 which contains 9323 bases, showing three single-nucleotide polymorphism (SNPs) in the beta globin gene region in chromosome number 11, besides the haplotypes that were appeared in the samples under investigation. This study also showed a significant correlation between SNP2 - SNP3 and between SNP 1 - SNP3, and a negative correlation between SNP1 - SNP2.

Conclusion: This study has used genome-wide association study (GWAS) in understanding the genetic diversity that explains the phenotypic shape of sickle cell disease (SCD) patients in Saudi Arabia. It is therefore important to conduct further studies at a large level in Saudi Arabia to confirm these important results, which will increase current understanding of the SCD's nature.

Keywords: Genetic origin, Mutation, Sickle cell disease, Single nucleotide, Single phenotype

INTRODUCTION

Sickle cell disease (SCD) is a major health problem according to the World Health Organization, and there are more than half million new cases each year¹⁻⁴. SCD is featured by several clinical signs such as: stroke, chest pain, abnormal hemoglobin in red blood cells, and recurrent vasoconstriction⁵. Chronic hemolysis usually leads to tuberculosis, delayed growth, anemia, and jaundice. Some SCD patients are sensitive for rheumatism, leg ulcers, pulmonary arterial hemorrhage and hypertension⁶⁻⁹. At the national level, SCD represents about 0.4-8% while SCD carriers ranged from 2% to 27%¹⁰. Currently, in Saudi Arabia, approximately 3 million individuals had genetic blood diseases in Saudi Arabia, of whom 30% lived in Dammam Region¹¹. Several publications showed that there are at least two different types of sickle cell anemia, the average type centralized in Dammam region,

while the severe type based in southwestern region¹²⁻¹⁹. Diseases such as cancer or SCD resulting from DNA variation or gene mutations can be investigated thoroughly via single nucleotide polymorphisms (SNPs) using a newly developed technique named as microarray, which depends on hybridization^{20,21}. The aim of this study was to conduct complete gene survey using microarray technology to investigate genetic pathogenicity of SCD.

MATERIALS AND METHODS

Patients: The study was conducted on 90 of sickle cell anemia patients selected randomly from the attending the blood diseases clinic at at two regional hospitals based in Riyadh, Saudi Arabia: The King Khalid University Hospital (KCUH), and The King Faisal Specialist Hospital

* Department of Family and Community, College of Medicine
** Department of Biology, College of Science,
Princess Nourah bint Abdulrahman University, Riyadh 11671, Saudi Arabia
*** Department of Immunology
College of Medicine, Um Qura University
**** Department of Medical Laboratory Science College of Applied Medical Science
Prince Sattam University
Saudi Arabia. E-mail: f.alenzi@psau.edu.sa
***** Department of Medical Laboratory Science
College of Applied Medical Sciences
Majmma University
***** Department of Medical Laboratory Science
Prince Sultan Military College for Health Sciences
***** Department of Dentistry, Riyadh Alm University
***** Al-Marafa University

Research Center (KFSHRC), from January 2017 to June 2020. The study protocol was processed according to the Declaration of Helsinki. Written informed consent from all patients were obtained. Ethical approval is obtained from both KCUH, and KFSHRC, Saudi Arabia.

Samples: Ten ml of venous blood was withdrawn from each patient and distributed to two tubes (each containing 5 ml) of ethylenediamine tetra acetic acid (EDTA). All samples were processed in the KCUH, pathology department. The Adevia 2120 (Siemens Company) was used for measuring hematology markers and Architect from Abbott Company for measuring biochemical markers.

Sequencing Analysis of PCR Results Using Biosystems Analyzer: Sequencing analysis refers to the determination of the nucleotide arrangement (G, A, T, and C) along with the DNA. It is an important and useful process in biotechnology, since it allows discovery of mutations and accurate diagnosis of genetic diseases using the 3730 DNA Analyzer from Applied Biosystems (DNA Sequencing by Capillary Electrophoresis Applied Biosystems Chemistry Guide Second Edition).

Microarray Technology: The protocol used in this study was applied as advised by the manufacture. GeneChip- Human Mapping 10K Array and Manual kit – Microarray Affymetrix (California, USA). Haploview, an important software program in bioinformatics, was used to analyze data contained in Hap Map, and to evaluate the quality of genetic data for a particular disease^{22,23}.

Statistical Analysis: The data obtained was subjected to a statistical analysis using Window Excel and SPSS v17 statistical tools. ANOVAS tests for multiple comparisons and significant analysis (p < 0.05) were carried out.

RESULTS

The study was conducted on 90 patients with sickle cell anemia from the outpatients clinic of blood diseases at three major regional hospitals in Saudi Arabia. A comprehensive database was created including the patient's name, patient history, and biochemical and blood biometrics tests. The patient's data was also studied to determine the disease severity, the frequency of clinical symptoms and pain. Genome-wide association study (GWAS) was also carried out to find genes/genomic regions that contribute to the final manifestation of a disease by measuring the association between the sites of single-nucleotide polymorphism (SNPs) Figure-1 shows the nature of the patients with sickle cell anemia. 80% of the patients were homozygous, 15% were gametes, and 5% had thalassemia.

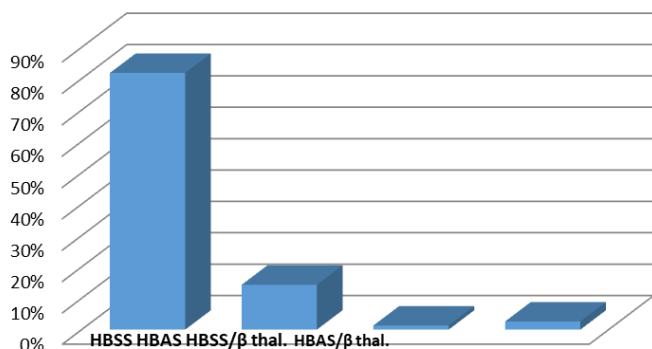


Figure 1: The nature of the patients

Clinical and Blood Data in the Research Sample: Table-1 presents a summary of hematological and biochemical measurements from

patients with sickle cell anemia that have been followed for more than 12 months. Table-2 also shows the statistical analysis of different parameters with their occurrence of in certain mutations.

Table 1: Hematological and biochemical measurements from SCA patients

	White Blood Cells (WBC)	Hemoglobin	Platelets	LDH	Bilirubin
Mean	11.5	95	371	440	51
Std. Deviation	5	23	152	201	32

Table 2: Relationship with changes found in beta-globin with P- Value

Parameter	Mutation	Significance Level (P- Value)
High WBC count	Mutated HBBF8_3	0.03
High Hemoglobin level	HBBF4_3	0.04
	HBBF5_1	0.05
High LDH level	HBBF8_2	0.02
	Mutated HBBF9_1	0.009
High the bilirubin level	Mutated HBBF9_1	0.003

Genome-wide association studies (GWAS) are used to find genes or genomic regions that contribute to diseases and genetic variation. In this study, the sites of a single-nucleotide polymorphism (SNPs) on SCD patients were examined. For this purpose, one of the most important programs in the field of bioinformatics was the haploview, which provides a comprehensive set of tools for analyzing large areas of the genome in many ways. In this study, genotype was performed using the AffymetrixChip technique. Three SNPs were found in beta region of chromosome 11. The haplotypes presented in the research samples are shown in Table 3 and Figures 2 and 3.

Table 3: Percentage of individual patterns of haplotypes found in the genotype of beta-globin for the samples

Percent	Haplotype	HapID
77.1%	AAA	Hap 1
12.7%	ACA	Hap 2
6.8%	ACC	Hap 3
3.4%	CCC	Hap 4

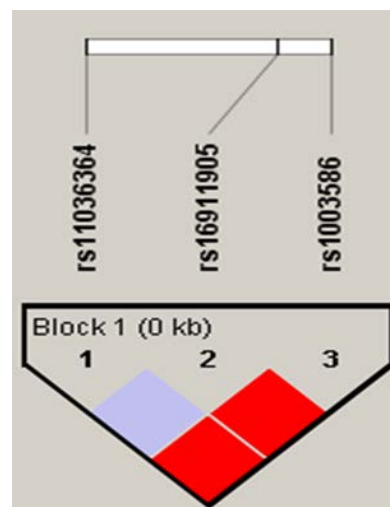


Figure 2: Single-nucleotide polymorphism (SNPs) identified in the beta region

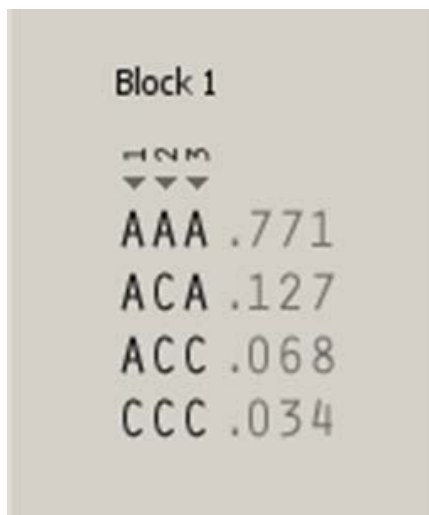


Figure 3: Individual Patterns Haplotypes that appeared in the beta gene-globin region of the study samples

DISCUSSION

The relationship between disease and genetic variation was examined using GWAS to determine the candidate genes that contribute to a particular disease. The SNPs are important markers that have been extensively examined in many correlative studies. Many studies have confirmed that these markers control a particular disease with different effects due to different genetic variants²⁴.

Important genetic links can be explained by a direct correlation, where SNP is associated with one or several of its multiple aspects. Therefore, the genetic association of the SNPs with the linkage disequilibrium (LD) determines the true causal variables. Another physiological study has confirmed the role of the presumed mutation in a disease²⁵.

In this study, three SNPs were identified in the beta-globin gene region of the SCD patients (SNP1: rs11036364), (SNP2: rs 16911905), and (SNP3: rs1003586). This study also showed a significant correlation between SNP2 - SNP3 and between SNP 1 - SNP3, and a negative correlation between SNP1 - SNP2. Previous study showed that SNPs in a single phenotype can be affected by the phenotype of the whole group²⁶.

SCD patients with the genotype AA had a higher risk of clinical distress than those with AC or CC pattern. The single nucleotide form detected in this study is different in its location and may be due to the differences in the genetic origin of our samples. A mutation in the beta-globin gene may be associated with a specific individual pattern.

Hattori et al²⁷ showed that on 98 SCD studies, 54% had Benin haplotype, while Bantu haplotype existed on 27%. Miller demonstrated that Arabian/Indian haplotype had 13% fetal hemoglobin²⁸, while African haplotype had lower HbF. It was implied that HbF had a clinical relevance²⁹. The SC mutation exists in Africa with several genetic haplotypes³⁰. Four haplotypes have been associated with HbS in Africa and the 5th is in India and/or Arabian Peninsula³¹ and are linked with disease severity³². These SCD haplotypes in Africa could increase the current understanding of genetic factors that shape the SCD phenotype. Currat et al study showed the exposure of a specific mutation (rs782144 SNP) across west African populations. They concluded that geographical distribution of known SC haplotypes is still not established in many African countries³³. Nogbri's group demonstrated that Arab Indian haplotype is one of the major HBB

haplotypes showing different clinical and hematological parameters compared to other haplotypes such as: Senegal, Cameroon and Benin³⁴.

CONCLUSION

In this study, linkage disequilibrium has been determined between single nucleotides polymorphism loci in the beta globin gene to identify which of them had a link to the disease appearance and affect. Promising data was obtained, however, larger and generalized samples from different locations from Saudi Arabia and Arab world are needed to confirm these results.

Authorship Contribution: All authors share equal effort contribution towards (1) substantial contributions to conception and design, acquisition, analysis and interpretation of data; (2) drafting the article and revising it critically for important intellectual content; and (3) final approval of the manuscript version to be published. Yes.

Potential Conflict of Interest: None.

Competing Interest: None.

Acceptance Date: 25 March 2021

Funding: This work was supported by the PSAU (grant no:2019/03/10211)

REFERENCES

1. WHO Report. Sickle-cell disease and other hemoglobin disorders. 2005-2011; Fact sheet N°308.
2. Weatherall DJ, Clegg JB. Inherited hemoglobin disorders: an increasing global health problem. *Bull World Health Organ* 2001;79(8):704-12.
3. Williams TN, Weatherall DJ. World distribution, population genetics, and health burden of the hemoglobinopathies. *Cold Spring Harb Perspect Med* 2012;2(9): a011692.
4. Henri, W, Kamran, M. Abnormal haemoglobins: detection & characterization. *Indian J Med Res* 2011;134(4):538-46.
5. Lionnet F, Hammoud N, Stojanovic KS. Hemoglobin sickle cell disease complications: a clinical study of 179 cases. *Haematologica* 2012;97(8):1136-41.
6. Bender MA, Hobbs W. Sickle Cell Disease. In: Pagon RA, Adam MP, Ardinger HH, Edited by GeneReviews (WA): University of Washington, Seattle 2003.
7. Whitley KS, Pace S. Sickle Cell Disease: A Phenotypic Patchwork. Imperial College Press, London. 2007.
8. Ballas SK, Gupta K, Adams-Graves P. Sickle cell pain: a critical reappraisal. *Blood* 2012;120(18):3647-56.
9. Abbas HA, Kahale M, Hosn MA. Pediatric Sickle Cell Disease. *Pediatr Ann* 2013; 42:3.
10. Jastaniah W. Epidemiology of sickle cell disease in Saudi Arabia. *Ann Saudi Med* 2011;31(3):289-93.
11. Al-Naseri E. Awareness of mothers in nutrition their children affected with sickle-cell anemia and beta-thalassemia in Jeddah. (Master Thesis). 2009, King Abdulaziz University, Jeddah, Kingdom of Saudi Arabia.
12. El-Hazmi MAF. Aspect of human hemoglobin and hemoglobin haemoglobinopathies in the Arabian Peninsula- studies at genetics and molecular level. 1979; KACST report between 1982-1992.
13. El-Hazmi MAF. Abnormal hemoglobin and allied disorders in the Middle East - Saudi Arabia. In: Distribution and evolution of hemoglobin and globin loci. Bowman JE, ed. 1983:239-49.

14. El-Hazmi MAF. Incidence and Frequency of Haemoglobinopathies and Thalassemia in the Northwest Sector of Arabia. *Saudi Med J* 1985; 6:149-62.
15. El-Hazmi MAF, Bahakim HM, Al-Swailem AM. The features of sickle cell disease in Saudi children. *J Trop Paedr* 1990;36(4):148-55.
16. El-Hazmi MAF, Jabbar FA, Al-Faleh FZ. Pattern for sickle cell, thalassemia and glucose 6 phosphate dehydrogenase deficiency genes in northwestern Saudi Arabia. *Hum Hered* 1991;41(1):26-34.
17. El-Hazmi MAF. Haemoglobinopathies, thalassemia and enzymopathies in Saudi Arabia. *Saudi Med J* 1992;13(6):488-99.
18. El-Hazmi MAF, Warsy AS. Appraisal of sickle cell and thalassemia genes in Saudi Arabia. *East Mediterr Health J* 1999;5(6):1147-53.
19. Al-Qurashi MM, El-Mouzan MI, Al-Herbish AS. The prevalence of sickle cell disease in Saudi children and adolescents. A community-based survey. *Saudi Med J* 2008; 29(10):1480-3.
20. Botstein D, Risch N. Discovering genotypes underlying human phenotypes: past successes for mendelian disease, future approaches for complex disease. *Nat Genet* 2003; 33:228-37.
21. Chanock, S. Candidate genes and single nucleotide polymorphisms (SNPs) in the study of human disease. *Dis Markers* 2001;17(2):89-98.
22. Barrett JC, Fry B, Maller J. Haploview: Analysis and visualization of LD and haplotype maps. *Bioinformatics* 2005;21(2):263-5.
23. Barrett JC. Haploview: Visualization and Analysis of SNP Genotype Data. *Cold Spring Harb Protoc* 2009:10.
24. Lohmueller KE, Mauney MM, Reich D. Variants associated with common disease are not unusually differentiated in frequency across populations. *Am J Hum Genet* 2006;78(1):130-6.
25. Lewis CM, Knight J. Introduction to genetic association studies. *Cold Spring Harb Protoc* 2012; 3:297-306.
26. Corona E, Chen R, Sikora M. Analysis of the genetic basis of disease in the context of worldwide human relationships and migration. *PLoS Genet* 2013;9(5):e1003447.
27. Hattori Y, Kutlar F, Kutlar A. Haplotypes of beta S chromosomes among patients with sickle cell anemia from Georgia. *Hemoglobin* 1986;10(6):623-42.
28. Miller BA, Salameh M, Ahmed M. Analysis of hemoglobin F production in Saudi Arabian families with sickle cell anemia. *Blood* 1987;70(3):716-20.
29. Nagel RL, Fabry ME, Pagnier J. Hematologically and genetically distinct forms of sickle cell anemia in Africa. The Senegal type and the Benin type. *N Engl J Med* 1985;312(14):880-4.
30. Labie D, Pagnier J, Lapoumeroulie C. Common haplotype dependency of high G gamma-globin gene expression and high Hb F levels in beta-thalassemia and sickle cell anemia patients. *Proc Natl Acad Sci USA*. 1985;82(7):2111-4.
31. Elion J, Berg PE, Lapoumeroulie C, et al. DNA sequence variation in a negative control region 5' to the beta-globin gene correlates with the phenotypic expression of the beta s mutation. *Blood* 1992;79(3):787-92.
32. Steinberg MH. Genetic etiologies for phenotypic diversity in sickle cell anemia. *Scientific World J* 2009;9:46-67.
33. Currat M, Trabuchet G, Rees D. Molecular analysis of the beta-globin gene cluster in the NiokholoMandenka population reveals a recent origin of the beta(S) Senegal mutation. *Am J Hum Genet* 2002;70(1):207-23.
34. Nongbri SRL, Verma HK, Lakkakula BV. Presence of atypical beta globin (HBB) gene cluster haplotypes in sickle cell anemia patients of India. *Rev Bras Hematol Hemoter* 2017;39(2):180-2.